

Novel Reversible Monoamine Oxidase A Inhibitors: Highly Potent and Selective 3-(1*H*-Pyrrol-3-yl)-2-oxazolidinonesSergio Valente,[†] Stefano Tomassi,[†] Giampiero Tempera,[‡] Stefania Saccoccio,[‡] Enzo Agostinelli,^{*,‡} and Antonello Mai^{*,†}[†]Dipartimento di Chimica e Tecnologie del Farmaco, Istituto Pasteur, Fondazione Cenci Bolognetti, Sapienza Università di Roma, Piazzale Aldo Moro 5, 00185 Roma, Italy[‡]Dipartimento di Scienze Biochimiche Rossi-Fanelli and Istituto di Biologia e Patologia Molecolare del CNR, Istituto Pasteur, Fondazione Cenci Bolognetti, Sapienza Università di Roma, Piazzale Aldo Moro 5, 00185 Roma, Italy

Supporting Information

ABSTRACT: Monoamine oxidases (MAOs) are involved in various psychiatric and neurodegenerative disorders; hence, MAO inhibitors are useful agents in the therapy of Parkinson's disease, Alzheimer's dementia, and depression syndrome. Herein we report a novel series of 3-(1*H*-pyrrol-3-yl)-2-oxazolidinones **3–7** as reversible, highly potent and selective anti-MAO-A agents. In particular, **4b**, **5b**, and **4c** showed a $K_{i-MAO-A}$ of 0.6, 0.8, and 1 nM, respectively, **4c** being 200000-fold selective for MAO-A with respect to MAO-B.

INTRODUCTION

Mitochondrial monoamine oxidases (MAOs, EC 1.4.3.4) are primarily involved in the metabolism of the biogenic monoamines. They catalyze the oxidative deamination of structurally different amines including the neurotransmitters dopamine, norepinephrine, serotonin (5-HT), tyramine, 2-phenethylamine (PEA), and exogenous arylalkylamines.^{1,2} The electron acceptor of the reaction is O₂, which is converted to H₂O₂ by the enzymes. MAOs are flavin adenine dinucleotide (FAD) containing enzymes located in the outer mitochondrial membrane of neuronal, glial, and other cells. Two different isoenzymatic forms have been identified, MAO-A and MAO-B, and are distinguishable by their differences in their amino acids sequences,³ three-dimensional structures,^{4–8} substrate specificity, and sensitivity to inhibitors.⁹ MAO-A preferentially catalyzes the oxidation of 5-HT and norepinephrine, and it is selectively inhibited by clorgyline, whereas MAO-B catalyzes the oxidation of PEA and benzylamine, and it is selectively inhibited by selegiline (1-deprenyl) and rasagiline.¹⁰ Tyramine and tryptamine appear to be substrates for both isoforms. Most human tissues express both isoenzyme, nevertheless blood platelets and myocardium are particularly rich in MAO-B, while high levels of MAO-A are detected in the human placenta, lung, and small intestine.¹¹

Isoenzyme A occurs in catecholaminergic neurons, whereas isoenzyme B is mainly present in neurons and glial cells of the human brain and also in other different cell types. The different localization suggests that the two subtypes have different physiological functions. In fact, MAO-A and -B are probably related to psychiatric and neurological disorders such as depression and Parkinson's disease (PD), respectively.¹² Several selective and reversible MAO-A inhibitors¹³ have been described as antidepressants (i.e., moclobemide, brofaromine, toloxatone, Chart S1 in Supporting Information), and selective and reversible/irreversible MAO-B inhibitors have been shown as useful in the treatment of PD (i.e., rasagiline, lazabemide,

safinamide, Chart S1). Recent reversible MAO-A inhibitors include amifuraline,¹⁴ a selective peripheral inhibitor, and CX157,¹⁵ specific for inhibition of MAO-A in human brain. Against MAO-B PF9601N,¹⁶ endowed with antioxidant/neuroprotective properties in an experimental model of PD, and M-30,¹⁷ a brain selective agent with antioxidant and neuroprotective properties for Alzheimer's disease, have been reported. In addition to neurological disorders, MAO-A seems to play a role in cardiac cellular degeneration¹⁸ and high expression of MAO-A in normal basal prostatic epithelium, and high-grade primary prostate cancer (PCa) has been identified. Moreover, clorgyline has been shown to inhibit several oncogenic pathways in PCa cells, suggesting MAO-A inhibitors as prodifferentiation and antioncogenic agents for high-risk PCa.¹⁹ Phenelzine directly and potentially inhibits adipocyte lipid storage and differentiation,²⁰ while selegiline was a potent inducer for bone marrow stromal cell differentiation into neuronal phenotype.²¹ Finally, MAO-A and -B are the major enzyme systems involved in vivo in the oxidative metabolism of xenobiotic amines, drugs in particular.²²

Toloxatone (Humoryl), characterized by the presence of the oxazolidin-2-one as a core structure, was the first reversible and selective MAO-A inhibitor introduced in the clinical practice as antidepressant agent. Between 2002 and 2005, we reported a new series of 3-(1*H*-pyrrol-1-yl)-2-oxazolidinones (**1**) active as reversible, potent, and MAO-A selective inhibitors^{23,24} (Figure 1) and the 3-(1*H*-pyrrol-2-yl)-2-oxazolidinone derivatives (**2**),²⁵ in which the 2-oxazolidinone nucleus was inserted at the C2 position of the pyrrole.

As a logical succession, we have now investigated the C3 position of the pyrrole, and we planned the synthesis of the 3-(1*H*-pyrrol-3-yl)-2-oxazolidinone derivatives **3–7**, shifting the 2-oxazolidinone nucleus at the pyrrole C3 position and introducing at the pyrrole N1 position three different alkyl

Received: September 11, 2011

Published: October 21, 2011

groups (Figure 1), obtaining compounds active up to sub-nanomolar concentration.

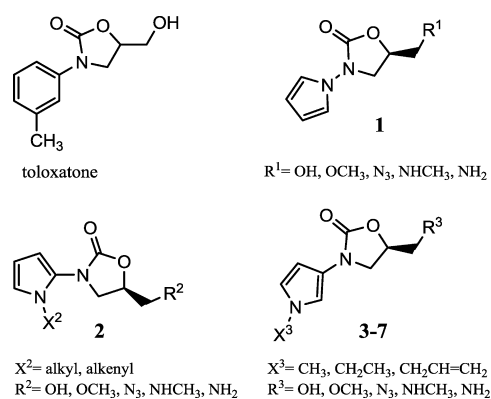


Figure 1. 1*H*-Pyrrolyl-2-oxazolidinones series.

CHEMISTRY

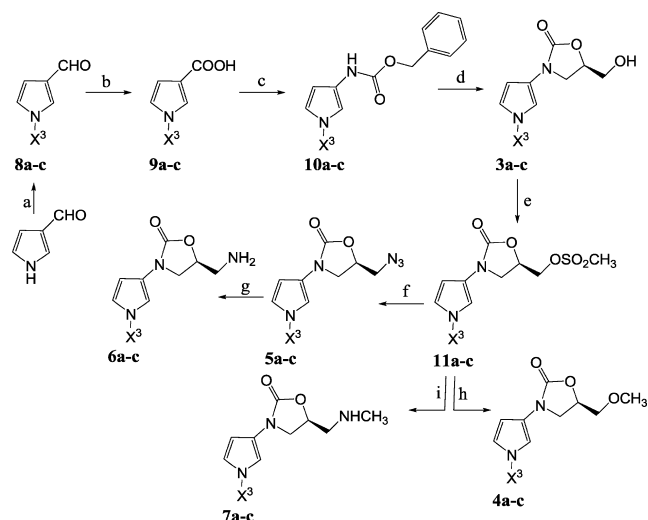
1-Alkyl-1*H*-pyrrole-3-carboxaldehydes **8a–c**, key intermediates for the synthesis of title compounds, were obtained by alkylation of 1*H*-pyrrole-3-carboxaldehyde²⁶ with the proper alkyl halide using potassium hydroxide as a base in dimethylsulfoxide. Afterward, 1-alkyl-1*H*-pyrrole-3-carboxylic acids **9a–c** were prepared by oxidation of the aldehydes **8a–c** with silver nitrate and 6 N sodium hydroxide in methanol or with potassium permanganate in water/2-propanone. After treatment with diphenylphosphoryl azide, triethylamine, and benzyl alcohol in benzene at 80 °C, the carboxylic acids **9a–c** afforded the 1-alkyl-3-benzyloxycarbonylamino-1*H*-pyrroles **10a–c** through Curtius rearrangement of the intermediate acylazides. Further reaction of **10a–c** with *n*-butyllithium in hexane at –78 °C followed by addition of (*R*)-glycidyl butyrate to the lithiated species furnished directly, after spontaneous hydrolysis of the butyrate function, the (*R*)-5-hydroxymethyl-3-(1-alkyl-1*H*-pyrrol-3-yl)-2-oxazolidinones **3a–c**.

The alcohols **3a–c** were then converted into the corresponding (*R*)-5-methanesulfonyloxymethyl derivatives **11a–c** with methanesulfonyl chloride and triethylamine, and the mesylates **11a–c** were subjected to nucleophilic displacement with sodium methoxide in methanol (room temperature) or with sodium azide in *N,N*-dimethylformamide (65 °C) or methylamine in tetrahydrofuran (65 °C) to afford the (*R*)-5-methoxymethyl-, the (*R*)-5-azidomethyl-, or the (*S*)-5-methylaminomethyl-3-(1-alkyl-1*H*-pyrrol-3-yl)-2-oxazolidinones **4a–c**, **5a–c**, or **7a–c**, respectively. From the (*R*)-5-azidomethyl derivatives **5a–c**, the corresponding aminomethyl compounds **6a–c** have been obtained by catalytic reduction (Scheme 1). Chemical and physical data for **3–7** and **8–11** are listed in Tables S1 and S2 of Supporting Information. Synthetic procedures for the synthesis of **3**, **4**, and **6** are reported below. Synthetic procedures for the synthesis of **5** and **7–11** are reported in Supporting Information.

RESULTS AND DISCUSSION

3-(1*H*-Pyrrol-3-yl)-2-oxazolidinones **3–7** have been tested against MAO-A and MAO-B enzymes, in comparison with toloxatone as a reference drug. Inhibitory data of befloxatone,²⁷ a more recent oxazolidinone anti-MAO, have been also added for comparison. Bovine brain mitochondria have been used as

Scheme 1^a



^a(a) (1) KOH, DMSO, room temp, 30 min; (2) X³-I or X³-Br, room temp, 1 h; (b) AgNO₃, NaOH, methanol, 70 °C or KMnO₄, 2-propanone/H₂O, room temp, overnight; (c) (1) (PhO)₂PON₃, Et₃N, benzene, 80 °C; (2) PhCH₂OH, benzene, 80 °C, overnight; (d) (1) *n*-BuLi, THF, N₂, –78 °C, 30 min; (2) (*R*)-glycidyl butyrate, N₂, 1 h, –78 °C, overnight at room temp; (e) CH₃SO₂Cl, Et₃N, CH₂Cl₂, 15 min; (f) NaN₃, DMF, N₂, 65 °C, overnight; (g) H₂, Pd/C, 1 h, room temp; (h) CH₃ONa, MeOH, N₂, room temp, overnight; (i) CH₃NH₂, THF, 80 °C, sealed tube, overnight.

the enzyme source and were isolated according to Basford.²⁸ Activities of MAO-A and MAO-B have been determined by a fluorimetric method with kynuramine as a substrate at several different concentrations.²⁹ The *K_i* values against the two MAO isozymes and the *A*-selectivity (expressed as *K_i*-MAO-B/*K_i*-MAO-A) are reported in Table 1. All tested compounds showed higher

Table 1. Monoamine Oxidase Inhibitory Activity of Compounds **3–7**^a

compd	X ³	R ³	<i>K_i</i> , μM		SI ^c
			MAO-A	MAO-B	
3a	methyl	OH	1	>10	>10
3b	ethyl	OH	0.003	0.3	100
3c	allyl	OH	0.35	625	1786
4a	methyl	OCH ₃	0.3	170	567
4b	ethyl	OCH ₃	0.0006	1.3	2167
4c	allyl	OCH ₃	0.001	200	200000
5a	methyl	N ₃	0.2	360	1800
5b	ethyl	N ₃	0.0008	20	25000
5c	allyl	N ₃	0.004	100	25000
6a	methyl	NH ₂	0.01	180	18000
6b	ethyl	NH ₂	0.13	25	192
6c	allyl	NH ₂	0.7	8	11
7a	methyl	NHCH ₃	0.007	>100	>14286
7b	ethyl	NHCH ₃	0.08	18	225
7c	allyl	NHCH ₃	0.1	2	20
toloxatone			0.38	15	40
befloxatone ^b			0.0025	0.22	88

^aData represent mean values of at least three separate experiments.

^bReference 27. ^cSelectivity index SI = *K_i*(MAO-B)/*K_i*(MAO-A).

anti-MAO-A than -MAO-B activity. Moreover, all derivatives displayed a reversible mode of action, since dialysis for 24 h in a

cold room against 0.1 M potassium phosphate buffer (pH 7.2) was able to restore 90–100% of the enzyme activity.

In the alcoholic series (3a–c), the MAO-A inhibitory activity depends on the substituent at the pyrrole N1 position. In fact, when we changed the methyl or allyl group with the ethyl one, the inhibiting activity shifted from micromolar (3a, $K_{i-MAO-A} = 1 \mu\text{M}$; 3c, $K_{i-MAO-A} = 0.35 \mu\text{M}$) to nanomolar level (3b, $K_{i-MAO-A} = 3 \text{ nM}$).

The 5-methoxymethyl derivatives 4a–c showed an anti-MAO-A activity increasing from low micromolar to sub-nanomolar range (4a, $K_{i-MAO-A} = 0.3 \mu\text{M}$; 4c, $K_{i-MAO-A} = 1 \text{ nM}$; 4b, 0.6 nM). Even in this series the ethyl group at the pyrrole N1 position was the most efficient substituent, despite the fact that the pyrrole N1-allyl derivative 4c showed the highest MAO-A selectivity ratio (200000).

Among the azides 5a–c, the 5-azidomethyl-3-(1-ethyl-1H-pyrrol-3-yl)-2-oxazolidinone (5b, $K_{i-MAO-A} = 0.8 \text{ nM}$) was almost as active as 4b against MAO-A, while its MAO-B inhibitory concentration is 25000-fold higher than MAO-B. This series showed an activity trend similar to that of the 5-hydroxymethyl and 5-methoxymethyl series (N1-substituents potency degree: ethyl > allyl > methyl). In the 5-aminomethyl series 6a–c, the compounds were endowed with a moderate inhibitory activity and MAO-A selectivity, they being active in the low micromolar range, with the exception of the pyrrole N1-methyl derivative 6a that showed $K_{i-MAO-A} = 10 \text{ nM}$ and SI = 18000.

Finally, the 5-methylaminomethyl compounds 7a–c showed an inversion of the reported structure–activity relationship: an increase in the size of the pyrrole N1-substituent produced less active and selective derivatives (7a, $K_{i-MAOA} = 7 \text{ nM}$, $K_{i-MAOB} > 100 \mu\text{M}$, SI ≥ 14286 ; 7c, $K_{i-MAOA} = 0.1 \mu\text{M}$, $K_{i-MAOB} = 2 \mu\text{M}$, SI = 20).

In conclusion, we have reported a novel series of 5-substituted 3-(1-alkyl-1H-pyrrol-3-yl)-2-oxazolidinones 3–7 as reversible, highly potent, and selective anti-MAO-A agents. Compared to the related 3-(1H-pyrrol-1-yl)- and 3-(1H-pyrrol-2-yl)-2-oxazolidinones 1 and 2,^{23,25} 3–7 were in general more efficient in inhibiting MAO-A: the most potent derivatives 4b and 5b are about 5-fold more effective than the best-scoring compounds of the 1 and 2 series, and the unique SI shown by 4c (200000) is about 20-fold higher than those displayed by the best compounds of 1 and 2. With respect to toloxatone, 4b, 4c, and 5b show 633-, 380-, and 475-fold higher MAO-A inhibitory activity, respectively. In comparison with befoxatone, 4b, 4c, and 5b present similar anti-MAO-A activity but increased A-selectivity ratio. Such results suggest that these compounds could be useful as novel promising antidepressant agents.

EXPERIMENTAL SECTION

Chemistry. Melting points were determined on a Buchi 530 melting point apparatus and are uncorrected. ¹H NMR and ¹³C NMR spectra were recorded at 400 MHz on a Bruker AC 400 spectrometer. Chemical shifts are reported in δ (ppm) units relative to the internal reference tetramethylsilane (Me₄Si). EIMS spectra were recorded with a Fisons Trio 1000 spectrometer; only molecular ions (M⁺) and base peaks are given. All compounds were routinely checked by TLC and ¹H NMR. TLC was performed on aluminum-backed silica gel plates (Merck DC, Alufolien Kiesegel 60 F254) with spots visualized by UV light. All solvents were reagent grade and, when necessary, were purified and dried by standard methods. Concentration of solutions after reactions and extractions involved the use of a rotary evaporator operating at reduced pressure of ~20 Torr. Organic solutions were dried over anhydrous sodium sulfate. Elemental analysis has been used

to determine purity of the described compounds, that is >95%. Analytical results are within $\pm 0.40\%$ of the theoretical values (see Table S3 in Supporting Information). All chemicals were purchased from Aldrich Chimica, Milan (Italy), or from Alfa Aesar, Milan (Italy), and were of the highest purity.

General Procedure for the Synthesis of (R)-5-Hydroxymethyl-3-(1-alkyl-1H-pyrrol-3-yl)-2-oxazolidinones (3a–c). Example: **Synthesis of (R)-5-Hydroxymethyl-3-(1-ethyl-1H-pyrrol-3-yl)-2-oxazolidinone (3b).** *n*-Butyllithium (2.5 M in hexane, 3.3 mmol, 1.32 mL) was added dropwise, over a period of 5 min, under nitrogen atmosphere at $-78 \text{ }^\circ\text{C}$ to a solution of 1-ethyl-1H-2-benzyloxycarbonylaminopyrrole 10b (3.24 mmol, 0.79 g) in dry THF (10 mL). The mixture was stirred at $-78 \text{ }^\circ\text{C}$ for 1 h. Then it was followed by addition of (R)-glycidyl butyrate (3.3 mmol, 0.47 mL). The resulting mixture was stirred initially at $-78 \text{ }^\circ\text{C}$ for 1 h, and then it was kept at room temperature overnight. Afterward, the reaction was quenched by addition of saturated ammonium chloride solution (100 mL) and extracted with ethyl acetate (3 \times 50 mL). The combined organic extracts were washed with water (100 mL) and brine (100 mL) and dried. The residue obtained upon evaporation of solvent was chromatographed over silica gel by eluting with ethyl acetate to give the alcohol 3b. ¹H NMR (CDCl₃) δ 1.42 (t, 3H, CH₂CH₃), 2.73 (bs, 1H, OH exchangeable with D₂O), 3.73–3.82 (m, 2H, NCH₂), 3.87–3.96 (m, 4H, CH₂OH and CH₂CH₃), 4.74 (m, 1H, OCH), 6.07 (m, 1H, pyrrole α proton), 6.56 (m, 1H, pyrrole β proton), 6.92 (m, 1H, pyrrole α proton) ppm. ¹³C NMR (CDCl₃) δ 15.60, 47.0, 47.80, 62.30, 84.80, 103.60, 117.10, 121.80, 122.80, 153.0 ppm. MS (EI), m/z [M]⁺: 210.1004.

General Procedure for the Synthesis of (R)-5-Methoxymethyl-3-(1-alkyl-1H-pyrrol-3-yl)-2-oxazolidinones (4a–c). Example: **Synthesis of (R)-5-Methoxymethyl-3-(1-ethyl-1H-pyrrol-3-yl)-2-oxazolidinone (4b).** A solution of sodium metal (3.43 mmol, 0.079 g atom) in methanol (5 mL) was added to (R)-5-methanesulfonyloxymethyl-3-(1-ethyl-1H-pyrrol-3-yl)-2-oxazolidinone 11b (0.86 mmol, 0.25 g) in methanol (5 mL), and the resulting mixture was stirred under nitrogen atmosphere at room temperature overnight. The reaction was quenched with water and extracted with ethyl acetate (3 \times 50 mL). The combined organic extracts were washed with water (100 mL) and brine (100 mL) and dried. The residue obtained upon evaporation of solvent was purified by column chromatography (silica gel, ethyl acetate/chloroform 1:1) to give the methoxymethyl derivative 4b as a pure oil. ¹H NMR (CDCl₃) δ 1.40 (t, 3H, CH₂CH₃), 3.42 (s, 1H, OCH₃), 3.59–3.61 (m, 2H, NCH₂ and CH₂O), 3.72–3.76 (m, 1H, NCH₂), 3.85–3.94 (m, 3H, CH₂CH₃ and CH₂O), 4.69–4.76 (m, 1H, OCH), 6.05 (m, 1H, pyrrole β proton), 6.54 (m, 1H, pyrrole α proton), 6.91 (m, 1H, pyrrole α proton). ¹³C NMR (CDCl₃) δ 15.60, 47.0, 52.20, 55.10, 103.60, 113.10, 117.10, 121.80, 122.80, 153.0 ppm. MS (EI), m/z [M]⁺: 210.1004.

General Procedure for the Synthesis of (S)-5-Aminomethyl-3-(1-alkyl-1H-pyrrol-3-yl)-2-oxazolidinones (6a–c). Example: **Synthesis of (S)-5-Aminomethyl-3-(1-ethyl-1H-pyrrol-3-yl)-2-oxazolidinone (6b).** A suspension of (R)-5-azidomethyl-3-(1-ethyl-1H-pyrrol-3-yl)-2-oxazolidinone 5b (2.38 mmol, 0.56 g), methanol (60 mL), and palladium on 10% carbon placed in a Parr apparatus was hydrogenated at 50 psi and 25 $^\circ\text{C}$ for 1 h. At last, palladium was filtered and methanol was evaporated to afford an oily residue that was chromatographed over silica gel by eluting with 9:1 chloroform/methanol to provide the amine derivative 6b as a pure oil. ¹H NMR (CDCl₃) δ 1.90 (bs, 2H, NH₂ exchangeable with D₂O), 2.83–2.96 (m, 2H, CH₂NH₂), 3.61–3.69 (m, 1H, NCH₂), 3.85–3.94 (m, 3H, NCH₂ and CH₂CH₃), 4.52–4.67 (m, 1H, OCH), 6.36 (m, 1H, pyrrole β proton), 6.86 (m, 1H, pyrrole α proton), 7.08–7.30 (m, 1H, pyrrole α proton) ppm. ¹³C NMR (CDCl₃) δ 15.60, 43.80, 47.0, 48.90, 87.0, 103.60, 117.1, 121.80, 122.80, 153.0 ppm. MS (EI), m/z [M]⁺: 209.1164.

Mitochondria Preparation. See Supporting Information.

Biochemical Assay. All chemicals were commercial reagents of analytical grade and were used without further purification. In all experiments, MAO activity of the beef brain mitochondria was determined by a sensitive fluorimetric method according to Matsumoto et al.,²⁹ using kynuramine as a substrate at four different

final concentrations ranging from 5 μM to 0.1 mM. In all assays, the incubation mixtures contained 0.1 mL of 0.25 M potassium phosphate buffer at pH 7.4, 30 μL of mitochondria (from a homogenate solution, 6 mg/mL), and drug solutions. Drug derivatives were dissolved in dimethylsulfoxide (DMSO), and then added to the reaction mixture, at final concentrations ranging from 0 to 10^{-3} μM . The solutions were preincubated for 30 min at 38 °C before adding the substrate and then incubated for other 30 min at the same temperature. The inhibitory activities on MAO-A and -B were separately determined after incubation of the mitochondrial fractions for 30 min at 38 °C in the presence of their specific inhibitors (1 μM L-deprenyl to estimate MAO A activity and 1 μM clorgyline to assay the B isoform). Addition of perchloric acid ended the reaction. The samples were then centrifuged at 10000g for 5 min, and the supernatant was added to 2.7 mL of 0.1 N NaOH. Fluorometric measurements were recorded with a Perkin-Elmer LS 50B spectrofluorimeter, at $\lambda_{\text{exc}} = 317$ nm and $\lambda_{\text{em}} = 393$ nm. The protein concentration was determined according to Goa. Dixon plots were used to estimate the inhibition constant (K_i) of the inhibitors. Data represent the mean of three or more experiments each performed in duplicate.

■ ASSOCIATED CONTENT

Supporting Information

Chemical and physical data for 3–11 and general procedures for the synthesis of 5 and 7–11. This material is available free of charge via the Internet at <http://pubs.acs.org>.

■ AUTHOR INFORMATION

Corresponding Author

*For E.A.: phone, +3906-4991-0838; fax, +3906-4440062; e-mail, enzo.agostinelli@uniroma1.it. For A.M.: phone, +3906-4991-3392; fax, +3906-49693268; e-mail, antonello.mai@uniroma1.it.

■ ACKNOWLEDGMENTS

This work was partially supported by the Italian MIUR (Ministero dell'Istruzione, dell'Università e della Ricerca), by Istituto Superiore di Sanità "Project Italy-USA", by Istituto Pasteur-Fondazione Cenci Bolognetti, and by funds from MIUR-PRIN (Cofin). Thanks are due to Fondazione "Enrico ed Enrica Sovena" for the scholarship given to S.S. for supporting her Ph.D.

■ ABBREVIATIONS USED

MAO, monoamine oxidase; 5-HT, 5-hydroxytryptamine; PEA, 2-phenethylamine; FAD, flavin adenine dinucleotide; ROS, reactive oxygen species; DNA, deoxyribonucleic acid; PCA, prostate cancer; MAOI, monoamine oxidase inhibitor; PD, Parkinson's disease; EDTA, ethylenediaminetetraacetic acid; PES, polyethersulfone

■ REFERENCES

- (1) Singer, T. P. Perspectives in MAO: past present and future. *J. Neural Transm. (Suppl.)* **1987**, *23*, 1–23.
- (2) O'Brien, E. M.; Tipton, K. Biochemistry and mechanism of action of monoamine oxidase A and B. *Neurol. Dis. Ther.* **1994**, *21*, 31–76.
- (3) Weyler, W.; Hsu, Y. P.; Breakefield, X. O. Biochemistry and genetics of monoamine oxidase. *O. Pharmacol. Ther.* **1990**, *47*, 391–417.
- (4) Binda, C.; Newton-Vinson, P.; Hubalek, F.; Edmondson, D. E.; Mattevi, A. Structure of human monoamine oxidase B, a drug target for a treatment of neurological disorders. *Nat. Struct. Biol.* **2002**, *9*, 22–26.

- (5) Binda, C.; Li, M.; Hubalek, F.; Restelli, N.; Edmondson, D. E.; Mattevi, A. Insights into the mode of inhibition of human mitochondrial monoamine oxidase B from high-resolution crystal structure. *Proc. Natl. Acad. Sci. U.S.A.* **2003**, *100*, 9750–9755.

- (6) Binda, C.; Hubalek, F.; Li, M.; Herzig, Y.; Sterling, J.; Edmondson, D. E.; Mattevi, A. Crystal structures of monoamine oxidase B in complex with four inhibitors of the *N*-propargylaminoindane class. *J. Med. Chem.* **2004**, *47*, 1767–1774.

- (7) De Colibus, L.; Li, M.; Binda, C.; Lustig, A.; Edmondson, D. E.; Mattevi, A. Three-dimensional structure of human monoamine oxidase A (MAO-A): relation to the structures of rat MAO-A and human MAO-B. *Proc. Natl. Acad. Sci. U.S.A.* **2005**, *102*, 12684–12689.

- (8) Binda, C.; Wang, J.; Pisani, L.; Caccia, C.; Carotti, A.; Salvati, P.; Edmondson, D. L.; Mattevi, A. Structures of human monoamine oxidase B complexes with selective non covalent inhibitors: safinamide and coumarin analogs. *J. Med. Chem.* **2007**, *50*, 5848–5852.

- (9) Grimsby, J.; Ian, N. C.; Neve, R.; Chen, K.; Shih, J. C. Tissue distribution of human monoamine oxidase-A and oxidase-B messenger-RNA. *J. Neurochem.* **1990**, *55*, 1166–1169.

- (10) Youdim, M. B.; Riederer, P. F. A review of mechanisms and role of monoamine oxidase inhibitors in Parkinson's disease. *Neurology* **2004**, *63* (Suppl. 2), S32.

- (11) Saura, J.; Nadal, E.; Van den Berg, B.; Vila, M.; Bombi, J. A.; Mahy, N. Localization of monoamine oxidases in human peripheral tissues. *Life Sci.* **1996**, *59*, 1341–1349.

- (12) Mallajosyula, J. K.; Kaur, D.; Chinta, S. J.; Rajagopalan, S.; Rane, A.; Nicholls, D. G.; Di Monte, D. A.; Macarthur, H.; Andersen, J. K. MAO B elevation in mouse brain astrocytes results in Parkinson's pathology. *PLoS One* **2008**, *3*:e, 1616.

- (13) Youdim, M. B.; Edmondson, D. E.; Tipton, K. F. The therapeutic potential of monoamine oxidase inhibitors. *Nat. Rev. Neurosci.* **2006**, *7*, 295–309.

- (14) Gentili, F.; Pizzinat, N.; Ordener, C.; Marchal-Victorien, S.; Maurel, A.; Hofmann, R.; Renard, R.; Delagrangue, P.; Pignini, M.; Parini, A.; Giannella, M. 3-[5-(4,5-Dihydro-1H-imidazol-2-yl)-furan-2-yl]phenylamine (amifuraline), a promising reversible and selective peripheral MAO-A inhibitor. *J. Med. Chem.* **2006**, *49*, 5578–5586.

- (15) Fowler, J. S.; Logan, J.; Azzaro, A. J.; Fielding, R. M.; Zhu, W.; Poshusta, A. K.; Burch, D.; Brand, B.; Free, J.; Asgharnejad, M.; Wang, G. J.; Telang, F.; Hubbard, B.; Jayne, M.; King, P.; Carter, P.; Carter, S.; Xu, Y.; Shea, C.; Muench, L.; Alexoff, D.; Shumay, E.; Schueller, M.; Warner, D.; Apelskog-Torres, K. Reversible inhibitors of monoamine oxidase-A (RIMAs): robust, reversible inhibition of human brain MAO-A by CX157. *Neuropsychopharmacology* **2010**, *35*, 623–31.

- (16) Perez, V.; Unzeta, M. PF 9601N [*N*-(2-propynyl)-2-(5-benzyloxy-indolyl)methylamine], a new MAO-B inhibitor, attenuates MPTP-induced depletion of striatal dopamine levels in C57/BL6 mice. *Neurochem. Int.* **2003**, *42*, 221–229.

- (17) Weinreb, O.; Mandel, S.; Bar-Am, O.; Amit, T. Iron-chelating backbone coupled with monoamine oxidase inhibitory moiety as novel pluripotential therapeutic agents for Alzheimer's disease: a tribute to Moussa Youdim. *J. Neural Transm.* **2011**, *118*, 479–92.

- (18) Bianchi, P.; Kunduzova, O.; Masini, E.; Cambon, C.; Bani, D.; Raimondi, L.; Seguelas, M. H.; Nistri, S.; Colucci, W.; Leducq, N.; Parini, A. Oxidative stress by mono amine oxidase mediates receptor-independent cardiomyocyte apoptosis by serotonin and postischemic myocardial injury. *Circulation* **2005**, *112*, 3297–3305.

- (19) Flamand, V.; Zhao, H.; Peehl, D. M. Targeting monoamine oxidase A in advanced prostate cancer. *J. Cancer Res. Clin. Oncol.* **2010**, *136*, 1761–1771.

- (20) Chiche, F.; Le Guillou, M.; Chetrite, G.; Lasnier, F.; Dugail, I.; Carpeno, C.; Moldes, M.; Feve, B. Antidepressant phenelzine alters differentiation of cultured human and mouse preadipocytes. *Mol. Pharmacol.* **2009**, *75*, 1052–1061.

- (21) Ghorbanian, M. T.; Tiraihi, T.; Mesbah-Namin, S. A.; Fathollahi, Y. Selegiline is an efficient and potent inducer for bone marrow stromal cell differentiation into neuronal phenotype. *Neurol. Res.* **2010**, *32*, 185–193.

(22) Strolin, B. M.; Tipton, K. F.; Whomsley, R.; Baltes, E. Factors affecting the relative importance of amine oxidases and monoxygenases in the in vivo metabolism of xenobiotic amines in humans. *J. Neural Transm.* **2007**, *114*, 787–791.

(23) Mai, A.; Artico, M.; Esposito, M.; Sbardella, G.; Massa, S.; Befani, O.; Turini, P.; Giovannini, V.; Mondovì, B. 3-(1*H*-Pyrrol-1-yl)-2-oxazolidinones as reversible, highly potent, and selective inhibitors of monoamine oxidase type A. *J. Med. Chem.* **2002**, *45*, 1180–1183.

(24) Mai, A.; Artico, M.; Valente, S.; Cerbara, I.; Befani, O.; Turini, P.; Dalla Vedova, L.; Agostinelli, E. Synthesis and biochemical evaluation of (*R*)-5-acyloxymethyl- and (*S*)-5-acylaminomethyl-3-(1*H*-pyrrol-1-yl)-2-oxazolidinones as new anti-monoamine oxidase (anti-MAO) agents. *ARKIVOC* **2004**, VT-941L, 32–43.

(25) Mai, A.; Artico, M.; Valente, S.; Sbardella, G.; Turini, P.; Befani, O.; Dalla Vedova, L.; Agostinelli, E. 3-(1*H*-Pyrrol-2-yl)-2-oxazolidinones as novel monoamine oxidase type A inhibitors. *Med. Chem.* **2005**, *1*, 117–124.

(26) Bray, B. L.; Mathies, P. H.; Naef, R.; Solas, D. R.; Tidwell, T. T.; Artis, D. R.; Muchowski, J. M. *N*-Triisopropylsilylpyrrole. A progenitor “par excellence” of 3-substituted pyrroles. *J. Org. Chem.* **1990**, *55*, 6317–6328.

(27) Rabasseda, X.; Sorbera, L. A.; Castaner, J. Bifloxadone. *Drugs Future* **1999**, *24*, 1057–1067.

(28) Stahl, W. L.; Smith, J. C.; Napolitano, L. M.; Basford, R. E. Brain mitochondria. I. Isolation of bovine brain mitochondria. *J. Cell Biol.* **1963**, *19*, 293–307.

(29) Matsumoto, T.; Suzuki, O.; Furuta, T.; Asai, M.; Kurokawa, Y.; Nimura, Y.; Katsumata, Y.; Takahashi, I. A sensitive fluorimetric assay for serum monoamine oxidase with kynuramine as substrate. *Clin. Biochem.* **1985**, *18*, 126–129.